

Water-Soluble Neutral Calix[4]arene–Lanthanide Complexes: Synthesis and Luminescence Properties

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Water-soluble calix[4]arenes **10a,b** with chromophores (“antenna”) attached to the lower rim via a short spacer are described. In the neutral lanthanide complexes of **10a,b** photoexcitation of the antenna induces lanthanide emission via intramolecular energy transfer. Calix[4]arene **10b** with a chrysene moiety as sensitizer shows strong lanthanide emission for Eu^{3+} with an excitation maximum at $\lambda = 363$ nm.

Introduction

There is considerable interest in the synthesis of molecular probes for imaging or bioassay purposes.^{1–3} Particularly interesting are lanthanide probes because of their unique characteristics like exceptionally large Stoke shifts (~ 200 – 300 nm), narrow emission line spectra, long-wavelength emission (~ 500 – 600 nm), and long luminescence lifetimes (~ 600 – 1000 ms).^{4,5} This explains the strong motivation for the synthesis of ligands for these lanthanide ions that form complexes with high kinetic stability and that are soluble in water for coupling reactions to biomolecules.^{6–13}

Various calix[4]arene derivatives are suitable for the complexation of trivalent cations, but in most cases they are not soluble in water.¹⁴ Only Sabbatini et al.¹⁵

reported the encapsulation of a lanthanide ion by a tetraamide calix[4]arene to give a water-soluble system, while Shinkai et al.¹⁶ prepared lanthanide complexes via complexation of the phenolic oxygen atoms of water-soluble calix[*n*]arenes. However, such lanthanide complexes based on calix[4]arenes are positively charged, which might lead to aspecific binding in biological systems.¹⁷

Recently, we described calixarene derivatives that are substituted with three carboxylic acid groups and which form *neutral* complexes with lanthanide cations.¹⁸ When we attached a triphenylene antenna chromophore to such calix[4]arenes they showed a strong sensitizing ability toward Eu^{3+} and Tb^{3+} , allowing the excitation of Eu^{3+} and Tb^{3+} with wavelengths extending to 350 nm.¹⁹

For a simple coupling to biomolecules, these calix[4]arene lanthanide probes should be water-soluble.²⁰ In the literature, different types of water-soluble calix[4]arenes have been reported that have hydrophilic substituents such as SO_3^- ,²¹ COO^- , PO_3^- ,²² $\text{SO}_2\text{N}(\text{CH}_2\text{CH}_2\text{OH})_2$,²³ N^+Me_3 ,²⁴ or polyethyleneoxy chains.^{25,26} Recently, we have reported neutral water-soluble calix[4]arenes by

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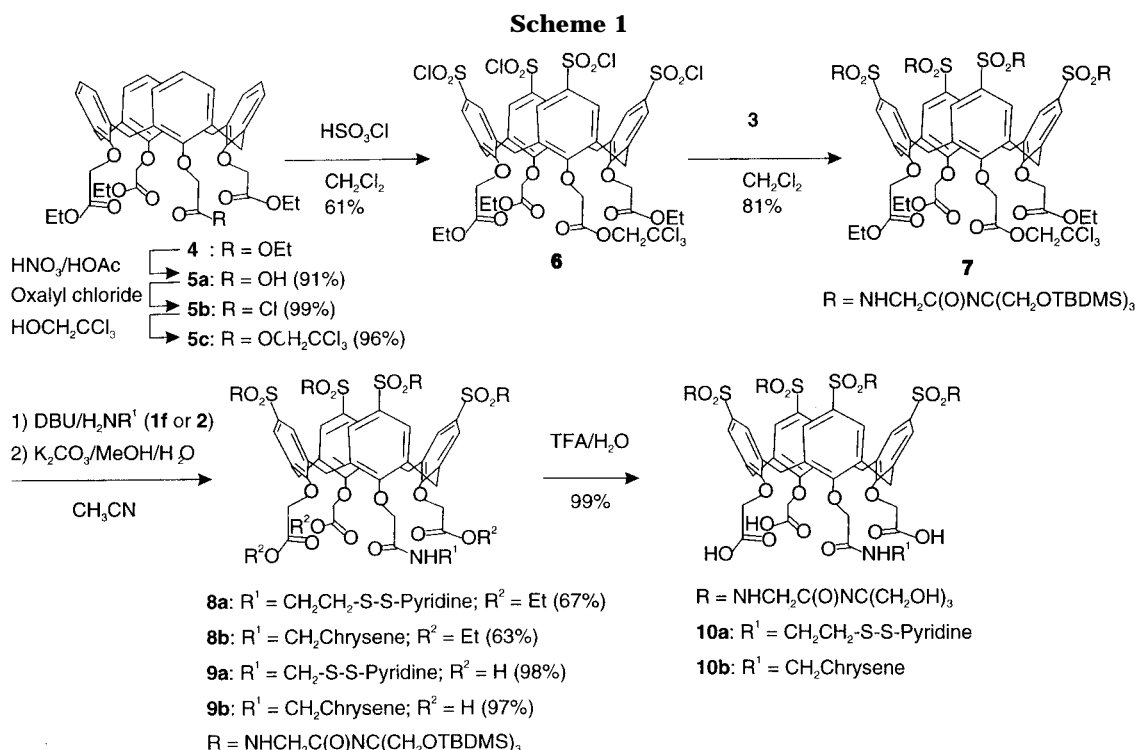
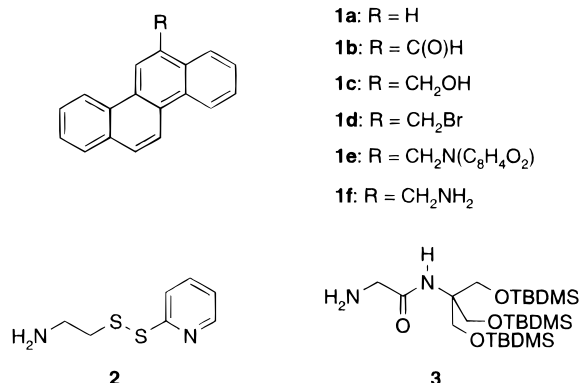
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**Chart 1**

the attachment of polyalcohol residues at the *upper rim* of the calix[4]arene.²⁷

In this paper, we report the synthesis and photophysical characterization of neutral, water-soluble Eu³⁺ and Tb³⁺ complexes substituted with two aromatic sensitizer groups (chrysene or 2-pyridyl disulfide).

Results and Discussion

Synthesis. The synthesis of water-soluble ionophores **10a,b** is depicted in Scheme 1. The water-solubility is introduced only in the last step of the synthesis because this facilitates handling and purification of the intermediates during the synthesis.

Reaction of triester-monoacid chloride calix[4]arene **5b**, prepared from the monocarboxylic acid **5a**,¹⁹ with 2,2,2-trichloroethanol²⁸ in CH₂Cl₂ gave the trichloroethyl ester **5c** in 96% yield. Reaction of trichloroethyl ester **5c** with chlorosulfonic acid in CHCl₃²⁹ afforded the tetrasulfonyl calix[4]arene **6** in 61% yield.³⁰ *N*-[Tris-

[[*tert*-butyldimethylsilyloxy]methyl]methane]-2-aminoacetamide (**3**)²⁷ (Chart 1) was coupled to calix[4]arene **6** to afford the tetrasulfonamide **7** in 82% yield. This intermediate can be used to introduce different amino-substituted residues. Substitution³¹ of the trichloroethyl group in **7** with 6-(aminomethyl)chrysene (**1f**) or *S*-(2-pyridylthio)cysteamine (**2**)³² (Chart 1) and DBU as a base in acetonitrile gave the corresponding amides **8a,b** in 63 and 67% yield, respectively. These sulfonamides (**8a,b**) were purified by chromatography over a NaCl-saturated silica column.³³

6-(Aminomethyl)chrysene (**1f**) was prepared from the known 6-(bromomethyl)chrysene³⁴ (**1d**) via the phthalimide **1e**. A Gabriel reaction of **1d** afforded the phthalimide **1e**, which after deprotection with hydrazine monohydrate gave the desired 6-(aminomethyl)chrysene (**1f**) in an overall yield of 95%. The precursor of **1d**, 6-(hydroxymethyl)chrysene (**1c**),³⁵ was obtained in 94% yield

(29) CHCl₃ was made ethanol free by passing it through an Al₂O₃ column in order to prevent the substitution of the trichloroethyl ester for an ethyl ester leading to the tetraethyl ester of tetrakis(chlorosulfonyl) calix[4]arene. To overcome the problem CH₂Cl₂ can also be used as a solvent.

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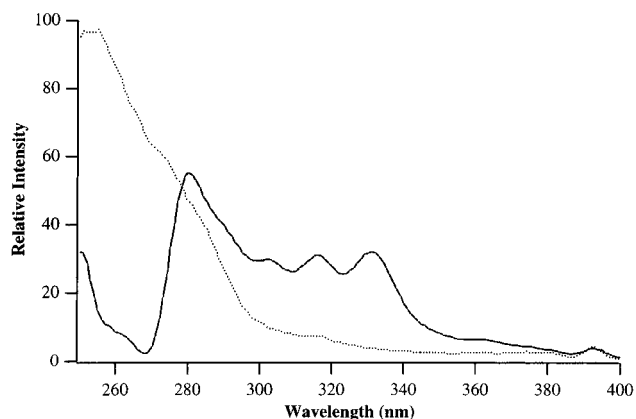


Figure 1. Corrected excitation spectra of Eu(**10a**) (···) and Eu(**10b**) (—) in Tris–HCl buffer (pH 8.0); $\lambda_{em} = 615$ nm. The spectra are normalized on the ${}^7F_0-{}^5L_6$ Eu³⁺ transition around 393 nm.

in an alternative way via reduction of 6-chrysenecarboxaldehyde (**1b**)³⁶ with BH₃·THF.

The selective hydrolysis of the three ethyl esters without cleavage of the amide bonds in **8a,b**, was achieved with K₂CO₃ in refluxing MeOH–H₂O–THF (5:1:2) to give the triacid derivatives **9a,b** in quantitative yield. Removal of the TBDMS-protective groups in **9a,b** with trifluoroacetic acid (TFA) gave the water-soluble derivatives **10a,b** in quantitative yield. Satisfactory mass spectra could be obtained with FRIT-FAB mass spectrometry. The ¹H NMR spectra of compounds **10a,b** are broad at room temperature but exhibit sharp signals at 80 °C, showing the characteristic AB system of the methylene bridges. This points to aggregation of the calix[4]arenes **10a,b** at room temperature.^{37,38} The maximum solubility of **10a** in water, determined by UV spectroscopy, is 8.0 mM at room temperature, and this is sufficient to couple **10a** to biomolecules.²⁰

Photophysical Properties. In order to investigate the sensitizing ability of **10a** and **10b** in aqueous solution, equivalent amounts of lanthanide ions (in the form of Eu(NO₃)₃·5H₂O or Tb(NO₃)₃·5H₂O) were added to a 1.5 × 10⁻⁴ M solution of **10a** or **10b** in Tris–HCl buffer pH 8.0. After deoxygenation by purging with argon for 15 min, the excitation spectra of these solutions were measured with the detection set at the emission wavelength of the lanthanide ion (615 nm for Eu³⁺ and 545 nm for Tb³⁺). The results for the Eu³⁺ complexes are shown in Figure 1. In this case, the excitation spectra in fact closely match the respective absorption spectra. Thus, efficient energy transfer occurs from both the 2-pyridyl disulfide and the chrysenes “antenna” leading to population of the Eu³⁺ emissive state. The chrysenes antenna in **10b** is particularly interesting because it allows excitation at wavelengths compatible with readily available light sources (e.g., a mercury arc) and with standard fluorescence microscope optics.

The spectral excitation region for compound Eu(**10b**) extends to the 0–0 transition of the chrysenes chro-

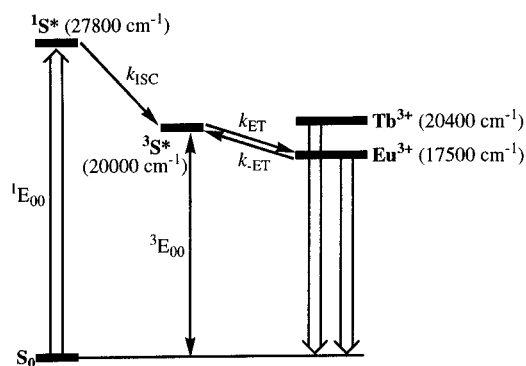


Figure 2. Energy scheme for the sensitized emission of Eu³⁺ and Tb³⁺ by chrysenes. Depicted are the main luminescent levels of Eu³⁺ (⁵D₀) and Tb³⁺ (⁵D₄).

mophore around 365 nm (see Figure 1). As we discussed before,¹⁹ this value must come close to the longest excitation wavelength possible for sensitization of Eu³⁺, since after excitation of the chromophore first intersystem crossing to the lowest triplet state is assumed to occur, which can subsequently give energy transfer to the luminescent lanthanide level (see Figure 2). For organic (poly)aromatic systems, a minimal single–triplet energy gap (¹E₀₀–³E₀₀) of 5000 cm⁻¹ may be assumed (in chrysenes this is 7800 cm⁻¹), while the triplet must still have high enough energy to make the energy transfer fast and to exclude possible (thermal) back-energy transfer. For chrysenes the triplet energy is 20 000 cm⁻¹,³⁹ only 2500 cm⁻¹ above the luminescent Eu³⁺ level, implying that in compound Eu(**10b**) probably already some back energy transfer occurs at room temperature.

Not unexpectedly, with Tb³⁺ it was found that **10b** does not allow sensitized luminescence to be observed. This again can readily be explained from the energy diagram depicted in Figure 2, assuming that energy transfer to the lanthanide ion occurs from the antenna triplet.

Interestingly, in the case of **10b** the effect of complexation with lanthanide ions is not restricted to quenching of the chrysenes triplet. This becomes evident from the effect of adding Eu(NO₃)₃·5H₂O to an aqueous solution of **10b** (8.5 × 10⁻⁵ M) on the total luminescence spectrum (see Figure 3a). In the absence of Eu³⁺, **10b** displays the typical fluorescence spectrum of the chrysenes unit ($\Phi_{fluor} = 0.15$, $\Phi_{ISC} = 0.85$ reported for chrysenes³⁹). Upon addition of Eu³⁺, the sensitized lanthanide emission appears, and its concentration dependence (see Figure 3b) nicely confirms formation of a 1:1 complex between **10b** and Eu³⁺. However, at the same time, the chrysenes fluorescence diminishes to about 10% of its original intensity. This fluorescence quenching seems to occur via nonspecific (dynamic) quenching and is not limited to a 1:1 stoichiometry. For the fluorescence quenching both direct energy transfer from the chrysenes singlet excited state to the lanthanide ion and enhanced intersystem crossing of chrysenes under the influence of a “heavy atom effect” of the lanthanide are viable mechanisms.⁴⁰ However, charge-transfer interaction between the singlet state of chrysenes and the lanthanide also may be involved. The excited-state oxidation potential of chrysenes is estimated to be ¹E_{ox}* = –2.1 V (calculated as the ground state oxidation potential E_{ox} = 1.35 V vs

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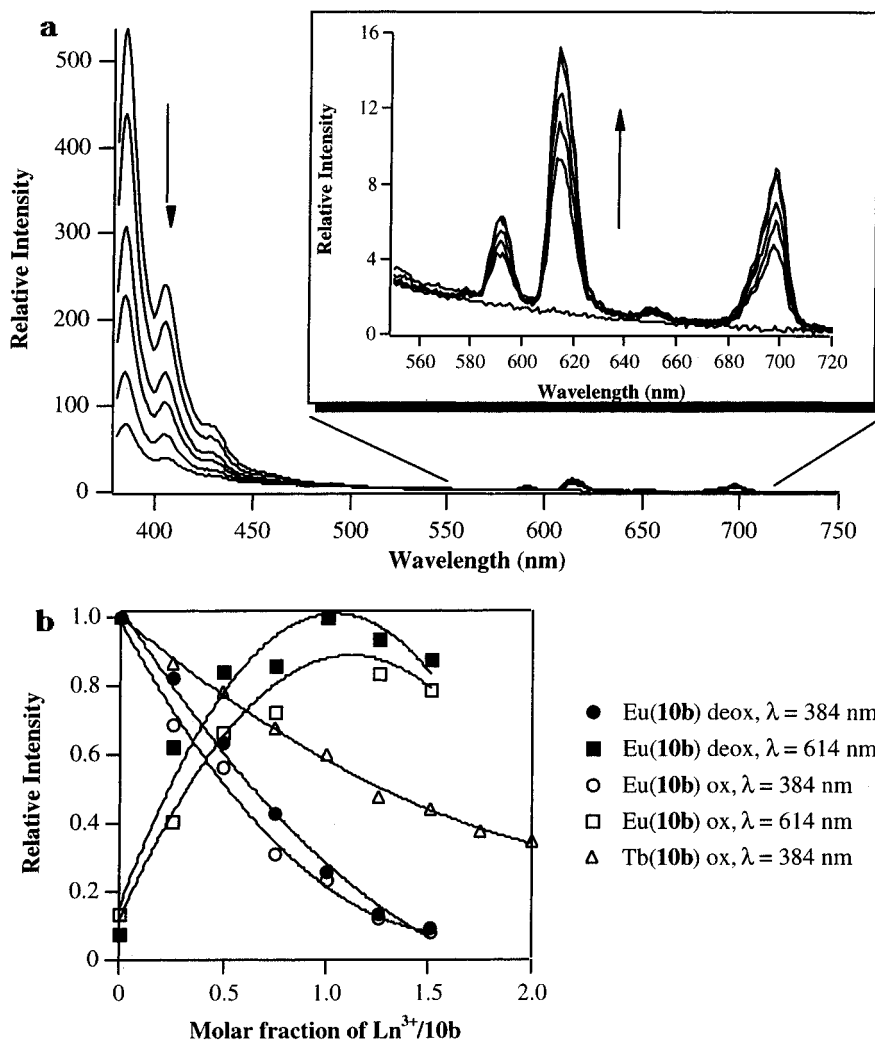


Figure 3. (a) Total luminescence spectra of **10b** (8.5×10^{-5} M) upon addition of increasing amounts of $\text{Eu}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$ in Tris-HCl buffer (pH 8.0); deoxygenated, $\lambda_{\text{ex}} = 363$ nm. (b) Relative luminescence intensities of the chrysenes fluorescence (384 nm) and Eu^{3+} luminescence (614 nm) upon addition of increasing amounts of $\text{Eu}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$ or $\text{Tb}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$ to **10b** in Tris-HCl buffer (pH 8.0) for both aerated (ox) and deoxygenated (deox) solutions; $\lambda_{\text{ex}} = 363$ nm.

SCE minus the 0–0 energy $E_{00} = 3.45$ V),³⁹ which allows electron transfer to Eu^{3+} ($E_{\text{red}} = -0.6$ V vs SCE)⁴¹ to occur. This conclusion is supported by the fact that the fluorescence of **10b** is quenched to a much lesser degree by Tb^{3+} (see Figure 3b). Tb^{3+} is very hard to reduce ($E_{\text{red}} = -3.9$ V vs SCE)⁴¹ and also very hard to oxidize ($E_{\text{ox}} = 2.9$ V vs SCE),⁴¹ making photoinduced electron transfer between Tb^{3+} and ¹chrysenes* impossible.

We have reported previously on sensitized luminescence of calix[4]arene/lanthanide complexes containing an antenna chromophore attached to the lower rim of the calixarene cage, to a complexed lanthanide ion, and employing three carboxylic acid groups as the complexing moieties, which resulted in overall neutral lanthanide complexes.^{18,19} Depending on the chromophore used, excitation wavelengths up to 350 nm could be achieved. However, these complexes could only be studied in organic solvents (e.g., methanol) because of their insolubility in aqueous media.

The present lanthanide complexes combine long-wavelength excitation (i.e., **10b**) and overall electroneutrality with water solubility via introduction of substituents at the upper rim of the calix[4]arene cage. The

luminescent lifetime for **Eu(10b)** is 0.11 and 0.14 ms in aerated and deoxygenated aqueous solutions, respectively. These lifetimes are shorter than those observed previously for calix[4]arene/ Eu^{3+} complexes in methanol (0.23–1.3 ms),^{18,19} thus demonstrating the stronger quenching ability of water. However, the present luminescence lifetimes are also certainly useful in schemes employing time-gated emission measurements to suppress the influence of the general background fluorescence of biological material, which usually has a lifetime in the nanosecond region.

While the 2-pyridyl disulfide group in **10a** only allows short wavelength excitation, it can be used as a coupling reagent toward SH functionalities in biological material.

Conclusions

We have developed a facile route to neutral, water-soluble calix[4]arene-based lanthanide complexes. Moreover, our approach has the potential for variation of substituents in the carboxamido fragment at the lower rim. Chrysenes gave high energy transfer from a 1:1 complex ligand/ Eu^{3+} at wavelengths extending to 363 nm. Currently, we are investigating the possibility of introducing both a sensitizer and a handle for coupling to biomolecules in the synthesis route.

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Experimental Section

Synthesis. General Procedure. Melting points are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃ with Me₄Si as the internal standard unless stated otherwise. Preparative column chromatography separations were performed on Merck silica gel 60 (230–400 mesh), while precoated silica gel plates (Merck, 60 F₂₅₄) were used for analytical TLC. FAB mass spectra were performed with *m*-nitrobenzyl alcohol as a matrix unless stated otherwise. CHCl₃, used in the chlorosulfonylation reaction, was flushed over an Al₂O₃ column and dried on molecular sieves (4 Å). All other chemicals were analytically pure and were used without further purification. Hexane refers to the fraction with bp 40–60 °C. All reactions were carried out under an argon atmosphere. Standard workup means that the organic layers were finally washed with water, dried over magnesium sulfate (MgSO₄), filtered, and concentrated *in vacuo*. The presence of solvents in the analytical samples was confirmed by ¹H NMR spectroscopy or potentiometric titrations. Fast atom bombardment (FAB) mass spectrometry for the water-soluble calix[4]arenes **10a,b** was carried out in a matrix solution (glycerol, thioglycerol, *m*-nitrobenzyl alcohol) onto a stainless steel probe and bombarded with xenon atoms with an energy of 3 KeV. During the high-resolution FRIT-FAB measurements a resolving power of 10,000 (10% valley definition) was used. Cesium iodide and/or glycerol were used to calibrate the mass spectrometer. *S*-(2-Pyridylthio)cysteamine hydrochloride,³² *de-tert*-butylated calix[4]arene, and calix[4]arene tetraethyl ester were prepared according to literature procedures.⁴²

Chrysene-6-carboxaldehyde (1b). To a solution of chrysene (**1a**) (2.0 g, 8.76 mmol) and freshly distilled TiCl₄ (3.30 g, 17.4 mmol) in dry CH₂Cl₂ (75 mL) was added dropwise bis-(chloromethyl) ether (10.0 g, 86.8 mmol) at –10 °C. The reaction mixture was stirred at rt for 30 min. The reaction was quenched by the addition of crushed ice (15 g). The organic layer was washed with 2 M HCl (2 × 100 mL), followed by standard workup. The crude reaction product was purified by flash column chromatography (CH₂Cl₂/hexanes, 50:50). The product was crystallized from CH₂Cl₂/hexane to obtain a yellow colored solid: yield 71%; mp 165–168 °C (lit.³⁶ mp 164–166 °C).

6-(Hydroxymethyl)chrysene (1c). To a solution of **1b** (1.0 g, 3.9 mmol) in dry THF (75 mL) was added BH₃·THF (3 mL, 1.0 M solution) at 0 °C. The cooling bath was removed, and the solution was stirred for 2 h at rt. HCl (1 M, 75 mL) was added, and the precipitate was filtered off. The product was triturated with CH₂Cl₂ to give pure **1c**: yield 94%; mp 200–203 °C (lit.³⁵ mp 200–202 °C).

6-(Phthalimidomethyl)chrysene (1e). To a solution of **1d** (0.50 g, 1.56 mmol) in DMF (25 mL) was added potassium phthalimide (0.32 g, 1.72 mmol). The solution was stirred for 5 h at 50 °C. The solvent was removed *in vacuo* and the residue dissolved in CH₂Cl₂ (50 mL) and washed with 0.1 M NaOH (3 × 50 mL). The product was obtained as a white solid in quantitative yield: mp 243–245 °C; ¹H NMR (DMSO-*d*₆) δ 9.1–9.0 (m, 1 H), 8.91 (d, 1 H, *J* = 8.4 Hz), 8.86 (s, 1 H), 8.84 (d, 1 H, *J* = 8.4 Hz), 8.5–8.4 (m, 1 H), 8.2–8.1 (m, 2 H), 8.0–7.86 (m, 4 H), 7.86–7.7 (m, 4 H); ¹³C NMR (DMSO-*d*₆) δ 166.28 (s), 42.69 (t); mass spectrum (EI) *m/z* 387.1 (M⁺, calcd for C₂₇H₁₇NO₂ 387.1).

6-(Aminomethyl)chrysene (1f). A solution of **1e** (0.50 g, 1.29 mmol) and hydrazine monohydrate (0.32 g, 6.46 mmol) in EtOH (50 mL) was refluxed for 2 h. The solvent was removed *in vacuo*, and H₂O (50 mL) and CH₂Cl₂ (50 mL) were added to the residue. The organic layer was washed with NaHCO₃ (saturated aqueous solution, 50 mL), followed by standard workup: yield 95%; mp 204–206 °C; ¹H NMR δ 8.84 (d, 2 H, *J* = 8.3 Hz), 8.73 (d, 1 H, *J* = 7.5 Hz), 8.70 (d, 1 H, *J* = 7.5 Hz), 8.22 (d, 1 H, *J* = 7.5 Hz), 8.00 (d, 2 H, *J* = 8.3 Hz), 7.8–7.5 (m, 4 H), 4.52 (s, 2 H), 1.69 (broad s, 2 H); ¹³C NMR δ 44.79; mass spectrum (EI) *m/z* 257.2 (M⁺, calcd 257.1). Anal. Calcd for C₁₉H₁₅N·0.4H₂O: C, 86.26; H, 5.72; N, 5.29. Found: C, 86.20; H, 5.49; N, 5.12.

25,26,27-Tris[(ethoxycarbonyl)methoxy]-28-[(hydroxycarbonyl)methoxy]calix[4]arene (5a) was prepared similarly to a literature procedure for the *p-tert*-butylcalix[4]arene analogue:¹⁹ white solid; yield 91%; mp 65–66 °C; ¹H NMR δ 7.13 (d, 4 H, *J* = 8.3 Hz), 7.0–6.8 (m, 2 H), 6.6–6.5 (m, 2 H), 6.5–6.4 (m, 4 H), 4.90 (d, 2 H, *J* = 15.0 Hz), 4.90, 4.85 (s, 4 H), 4.8–4.6 (m, 4 H), 4.40 (d, 2 H, *J* = 15.9 Hz), 4.3–4.2 (m, 6 H), 3.33 (d, 2 H, *J* = 12.5 Hz), 3.30 (d, 2 H, *J* = 12.5 Hz), 1.31 (t, 9 H, *J* = 8.3 Hz); FAB mass spectrum *m/z* 764.0 ([M + Na]⁺, calcd 764.3), *m/z* 739.5 (M⁺, calcd 739.3). Anal. Calcd for C₄₂H₄₄O₁₂: C, 68.10; H, 5.99. Found: C, 67.80; H, 5.95.

25,26,27-Tris[(ethoxycarbonyl)methoxy]-28-[[2,2,2-trichloroethoxy]carbonyl]methoxycalix[4]arene (5c). To a solution of triester monoacid **5a** (4.49 g, 6.9 mmol) in dry CH₂Cl₂ (75 mL) was added oxalyl chloride (5 mL), and the resulting reaction mixture was refluxed for 1 h. The solvent and excess of oxalyl chloride were removed *in vacuo*, whereupon 2,2,2-trichloroethanol (1.05 g, 7.0 mmol) and triethylamine (2.1 g, 20.8 mmol) were added. The mixture was stirred overnight at rt. Water (75 mL) was added, followed by standard workup. The crude product was separated by column chromatography (CH₂Cl₂/hexane 3/1) to give **5c** as a white solid: yield 96%; mp 124–125 °C; ¹H NMR δ 6.7–6.5 (m, 6 H), 6.47 (s, 6 H), 4.90 (s, 2 H), 4.89–4.65 (m, 4 H), 4.75, 4.70 (s, 4 H), 4.61 (d, 4 H, *J* = 6.7 Hz), 4.2–4.0 (m, 6 H), 3.18 (d, 2 H, *J* = 12.5 Hz), 3.17 (d, 2 H, *J* = 12.5 Hz), 1.3–1.2 (m, 9 H); FAB mass spectrum *m/z* 895.1 ([M + Na]⁺, calcd 895.2). Anal. Calcd for C₄₄H₄₅Cl₃O₁₂: C, 60.59; H, 5.20. Found: C, 60.51; H, 5.19.

5,11,17,23-Tetrakis(chlorosulfonyl)-25,26,27-tris-[(ethoxycarbonyl)methoxy]-28-[[2,2,2-trichloroethoxy]carbonyl]methoxycalix[4]arene (6). To a solution of trichloroethyl ester calix[4]arene **5c** (5.0 g, 5.7 mmol) in CH₂Cl₂ (100 mL) was added dropwise HSO₃Cl (18.7 mL, 0.23 mol) at –10 °C. When the addition was complete, the cooling bath was removed, and the reaction mixture was stirred at rt for 3 h. The solution was poured into a separation funnel filled with crushed ice (50 g). To the white suspension formed was added CH₂Cl₂ (100 mL), and the layers were allowed to separate for 24 h. The organic layer was concentrated, and the residue was purified by flash column chromatography (CH₂Cl₂/ethyl acetate (or THF), 90/10) to afford pure **6** as a white solid: yield 81%; mp 95–97 °C; ¹H NMR δ 7.60 (s, 2 H), 7.59 (s, 2 H), 7.27 (s, 4 H), 5.15 (d, 2 H, *J* = 12.5 Hz), 5.11 (d, 2 H, *J* = 12.5 Hz), 4.92 (s, 2 H), 4.81 (s, 6 H), 4.5–4.1 (m, 8 H), 3.55 (d, 2 H, *J* = 12.5 Hz), 3.52 (d, 2 H, *J* = 12.5 Hz), 1.4–1.2 (m, 9 H); FAB mass spectrum *m/z* 1289.4 ([M + Na]⁺, calcd 1289.2). Anal. Calcd for C₄₄H₄₁Cl₇O₂₀S₄: C, 41.74; H, 3.26. Found: C, 41.82; H, 3.21.

5,11,17,23-Tetrakis[[[[[tris[[[*tert*-butyldimethylsilyloxy]methyl]methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(ethoxycarbonyl)methoxy]-28-[[2,2,2-trichloroethoxy]carbonyl]methoxycalix[4]arene (7). To a solution of tetrasulfonyl calix[4]arene **6** (1.62 g, 1.3 mmol) was added dropwise a mixture of amine **3** (3.30 g, 6.3 mmol) and triethylamine (0.92 g, 9.1 mmol). The solution was stirred overnight. The solution was washed with 1 M HCl (50 mL) followed by standard workup. The crude solid was purified by column chromatography (CH₂Cl₂/ethyl acetate 97/3) to give **7** as a white solid: yield 79%; mp 102–104 °C; ¹H NMR δ 7.36 (broad s, 4 H), 6.72 (broad s, 2 H), 6.65 (broad s, 2 H), 6.48 (broad s, 4 H), 5.05 (broad s, 2 H) 5.0–4.6 (m, 12 H), 4.18 (q, 6 H, *J* = 6.7 Hz), 3.81 (broad s, 24 H), 3.4–3.1 (m, 12 H), 1.3–1.2 (m, 9 H), 0.83 (s, 108 H), 0.00 (s, 72 H); FAB mass spectrum *m/z* 3226.0 ([M + Na]⁺, calcd 3226.4). Anal. Calcd for C₁₄₀H₂₆₁Cl₃N₈O₃₆S₄Si₁₂·2H₂O: C, 52.16; H, 8.24; N, 3.48. Found: C, 52.02; H, 8.21; N, 3.45.

5,11,17,23-Tetrakis[[[[[tris[[[*tert*-butyldimethylsilyloxy]methyl]methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(ethoxycarbonyl)methoxy]-28-[[2-(pyridinedithio)ethyl]amino]carbonyl]methoxycalix[4]arene (8a). A solution of **7** (0.65 g, 0.2 mmol), *S*-(2-pyridylthio)cysteamine (**2**) (0.17 g, 0.2 mmol), triethylamine (0.03 g, 0.2 mmol), and DBU (0.28 g, 1.8 mmol) in acetonitrile (50 mL) was heated at 40 °C for 30 min. After removal of the

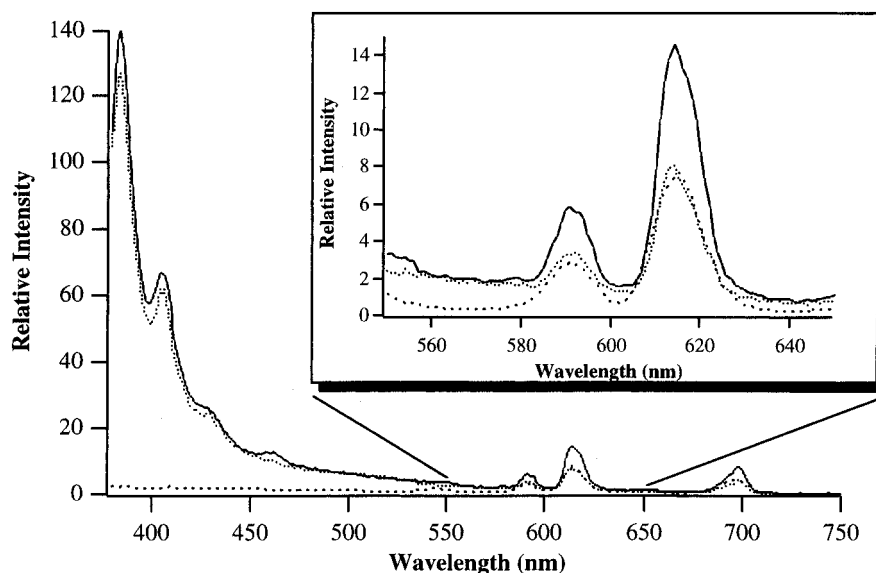


Figure 4. Corrected emission spectra of Eu(**10a**) (---; $\lambda_{\text{ex}} = 250$ nm) and Eu(**10b**) (aerated: ···; deoxygenated: —; $\lambda_{\text{ex}} = 363$ nm) in Tris-HCl buffer (pH 8.0) at room temperature.

solvent, the residue was dissolved in CH_2Cl_2 (50 mL) and the solution washed with 1 M HCl (50 mL), followed by standard workup. After column chromatography (silica gel saturated with NaCl, ethyl acetate), the product was obtained as a slightly yellow solid: yield 67%; mp 95–98 °C; $^1\text{H NMR}$ δ 8.43 (t, 1 H, $J = 4.2$ Hz), 8.05 (broad s, 1 H), 7.7–7.5 (m, 3 H), 7.4–7.1 (m, 8 H), 6.69 (broad s, 4 H), 6.58 (broad s, 4H), 5.0–4.6 (m, 16 H), 4.22 (q, 6 H, $J = 8.3$ Hz), 3.81 (s, 24 H), 3.69 (m, 2 H), 3.4–3.1 (m, 12 H), 2.99 (t, 2 H, $J = 6.7$ Hz), 1.3–1.2 (m, 9 H), 0.83 (s, 108 H), 0.00 (s, 72 H); FAB mass spectrum (ONPOE) m/z 3238.0 (M^+ , calcd 3237.5), 3128.6 ($[\text{M}^+ - \text{S-pyridine}]$). Anal. Calcd for $\text{C}_{145}\text{H}_{268}\text{N}_{10}\text{O}_{35}\text{S}_6\text{Si}_{12}$: C, 53.44; H, 8.36; N, 4.30. Found: C, 53.30; H, 8.51; N, 4.17.

5,11,17,23-Tetrakis[[[[[tris[[[(*tert*-butyldimethylsilyloxy)methyl]methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(ethoxycarbonyl)methoxy]-28-[[[9-chrysenemethyl]amino]carbonyl]methoxy]calix[4]arene (**8b**)] was prepared similarly as compound **8a** starting from the trichloroethyl ester **7** (1.65 g, 0.5 mmol), 6-(aminomethyl)chryseno (**1f**) (0.15 g, 0.6 mmol) and DBU (0.71 g, 4.6 mmol). The crude solid was purified by column chromatography (silica gel saturated with NaCl, ethyl acetate/ CH_2Cl_2 5/95): yield 63%; mp 115–117 °C; $^1\text{H NMR}$ δ 8.8–8.7 (m, 3 H), 8.66 (d, 1 H, $J = 9.3$ Hz), 8.17 (d, 1 H, $J = 7.5$ Hz), 8.09 (broad s, 1 H), 8.0–7.9 (m, 2 H), 7.8–7.5 (m, 4 H), 7.4–7.2 (m, 4 H coincides with CDCl_3), 6.8–6.5 (m, 8 H), 6.40 (broad s, 4 H), 5.20 (d, 2 H, $J = 6.2$ Hz), 4.9–4.4 (m, 12 H), 4.07 (q, 6 H, $J = 7.2$ Hz), 3.81 (broad s, 24 H), 3.4–3.2 (broad s, 12 H), 1.08 (t, 9 H, $J = 7.1$ Hz), 0.85 (s, 108 H), 0.01 (s, 72 H); FAB mass spectrum m/z 3333.6 ($[\text{M} - \text{H}]^+$, calcd 3333.6). Anal. Calcd for $\text{C}_{157}\text{H}_{273}\text{N}_9\text{O}_{35}\text{S}_4\text{Si}_{12} \cdot 2\text{H}_2\text{O}$: C, 56.32; H, 8.34; N, 3.76. Found: C, 55.93; H, 8.41; N, 3.58.

General Procedure for the Preparation of Triacid Calix[4]arenes 9a,b. To a solution of **8a,b** (5 mmol) in $\text{MeOH}/\text{H}_2\text{O}$ 5/1 (50 mL) and a minimum amount of THF, for solubility reasons, was added K_2CO_3 (50 mmol). After the solution was refluxed for 15 min, CH_2Cl_2 (50 mL) was added, and the solution was acidified with 1 M HCl to pH 7. The aqueous layer was extracted with CH_2Cl_2 (2 \times 50 mL). The combined layers were dried over MgSO_4 , filtered, and concentrated *in vacuo* to give the pure products.

The characterization of the derivatives **9a,b** with $^1\text{H NMR}$ spectroscopy was difficult because of line broadening. Additionally, the relative intensity of the ^1H CH_3 -resonances (TBDMS) compared to the other ^1H -resonances is large. No satisfactory mass spectrum (FAB and FRIT-FAB) could be obtained for the triacid derivatives **9a,b**. Therefore, characterization was done by melting point determination and elemental analysis.

5,11,17,23-Tetrakis[[[[[tris[[[(*tert*-butyldimethylsilyloxy)methyl]methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(hydroxycarbonyl)methoxy]-28-[[[2'-(pyridinedithio)ethyl]amino]carbonyl]methoxy]calix[4]arene (**9a**)] yield 98%; mp 143–144 °C. Anal. Calcd for $\text{C}_{139}\text{H}_{256}\text{N}_{10}\text{O}_{35}\text{S}_6\text{Si}_{12}$: C, 52.89; H, 8.17; N, 4.44. Found: C, 52.87; H, 8.34; N, 4.30.

5,11,17,23-Tetrakis[[[[[tris[[[(*tert*-butyldimethylsilyloxy)methyl]methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(hydroxycarbonyl)methoxy]-28-[[[9-chrysenemethyl]amino]carbonyl]methoxy]calix[4]arene (**9b**)] yield 97%; mp 160–162 °C. Anal. Calcd for $\text{C}_{151}\text{H}_{261}\text{N}_9\text{O}_{35}\text{S}_4\text{Si}_{12} \cdot 3.5\text{H}_2\text{O}$: C, 55.10; H, 8.11; N, 3.83. Found: C, 54.80; H, 8.10; N, 3.72.

General Procedure for the Preparation of Water-Soluble Calix[4]arenes 10a,b. A solution of triacid calix[4]arenes **9a,b** (0.5 g) in trifluoroacetic acid (TFA)/ H_2O (9/1, 25 mL) was stirred overnight, whereupon toluene (50 mL) was added. The solution was concentrated *in vacuo*. Again, toluene (50 mL) was added, and the mixture was concentrated *in vacuo*. This procedure was repeated three times. The crude white solid was dissolved in a minimum amount of H_2O (5 mL). To this solution was added dropwise acetonitrile (**10a**) or acetone (**10b**) until the product started to precipitate. After the suspension was stirred for 30 min, the product was filtered. The product was freeze-dried and analyzed. Prior to the luminescence measurements, solutions of compounds **10a,b** in water were dialyzed (benzoylated cellulose membrane) to remove excess trifluoroacetic acid and again freeze-dried.

5,11,17,23-Tetrakis[[[[[tris[(hydroxymethyl)methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(hydroxycarbonyl)methoxy]-28-[[[2'-(pyridinedithio)ethyl]amino]carbonyl]methoxy]calix[4]arene (**10a**)]. The crude solid was triturated with $\text{EtOAc}/\text{H}_2\text{O}$ to obtain pure **10a**: yield 99%; during melting point determination the compound decomposed; $^1\text{H NMR}$ (400 MHz, D_2O , 50 °C) δ 8.57 (s, 1 H), 8.0–7.9 (m, 3 H), 7.8–7.2 (m, 8 H), 5.4–4.9 (m, 12 H), 4.6–4.4 (m, 8 H), 4.2–3.6 (m, 24 H), 3.40 (t, 2 H, $J = 7.4$ Hz), 3.28 (d, 4 H, $J = 11.4$ Hz), 1.50 (t, 2 H, $J = 7.4$ Hz); FAB mass spectrum (glycerol) m/z 1785.7 (M^+ , calcd 1785.8). Anal. Calcd for $\text{C}_{67}\text{H}_{88}\text{N}_{10}\text{O}_{35}\text{S}_6 \cdot 3\text{TFA}$: C, 41.40; H, 5.20; N, 6.20. Found: C, 41.27; H, 5.05; N, 6.31.

5,11,17,23-Tetrakis[[[[[tris[(hydroxymethyl)methyl]amino]carbonyl]methyl]amino]sulfonyl]-25,26,27-tris[(hydroxycarbonyl)methoxy]-28-[[[9-chrysenemethyl]amino]carbonyl]methoxy]calix[4]arene (**10b**)]. The crude solid was triturated with ethyl acetate/ H_2O to obtain **10b** as a white pure solid: yield 99%; during melting point determination the compound decomposed; $^1\text{H NMR}$ (D_2O , 80 °C) δ 9.0–

7.2 (m, 19 H), 5.4–4.8 (m, 18 H), 4.69 (broad s, 24 H), 3.82 (d, 4 H, $J = 9.3$ Hz), 3.27 (d, 4 H, $J = 11.5$ Hz); FAB mass spectrum (glycerol) m/z 1857.8 [(M + H)⁺, calcd 1856.8]; exact FRIT-FAB m/z 1856.4735 [M⁺, calcd 1856.4761], the fragmentation pattern corresponds with the proposed structure. Anal. Calcd for C₇₉H₉₃N₉O₃₅S₄·4TFA: C, 45.10; H, 4.01; N, 5.43. Found: C, 45.18; H, 4.23; N, 5.45.

Luminescence Measurements. Continuous emission and excitation spectra were recorded on a Spex Fluorolog 2 spectrofluorimeter. Figure 4 gives representative results of such spectra.

Time-resolved emission spectra were obtained using a Lumonics EX700 XeCl excimer laser (308 nm) as excitation source. The resulting luminescence was observed by a gated, intensified CCD camera from Princeton Instruments. Spectra were taken with a typical gatewidth of 25 μ s, an initial delay

of about 0.5 μ s relative to the laser pulse maximum, and 50 μ s increment delay between spectra. At each delay, spectra were averaged over 50 laser shots to improve the signal to noise ratio. From these data, luminescent lifetimes were calculated by fitting the wavelength integrated signal in time. Monoexponential decay was observed in all cases.

All measurements were done in Tris–HCl buffer (0.05 M, pH 8.0).

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